



 UCSD

*Center for NMR Spectroscopy  
and Imaging of Proteins*



**Tutorial:**  
**Basic calibrations for solid state NMR  
experiments of membrane proteins**

*BTRC, 2014*

## *Useful calibrations in protein solid state NMR*

1. Referencing  $^1\text{H}$ ,  $^{15}\text{N}$ ,  $^{13}\text{C}$  and  $^{31}\text{P}$
2. Basic VT calibration
3. Determination of frictional heating inside an MAS sample
4. Determination of power deposition on an NMR sample (static or MAS experiments)

## 1. Useful samples for referencing

- $^{15}\text{N}$  ammonium sulfate, i.e., AMS (external reference). Set  $^{15}\text{N}$  resonance at 26.8 ppm.
- Adamantane, natural abundance (external reference). Set  $^{13}\text{C}$  high-frequency methylene resonance at 38.5 ppm
- DSS in deuterated water. Set the DSS resonance to 0 ppm, use to verify  $^1\text{H}$  chemical shift of HDO (internal reference for water) and, indirectly,  $^{13}\text{C}$  chemical shifts ( $\nu_0^{\text{C}} = 0.251449530 \cdot \nu_0^{\text{H}}$ ).
- $\text{H}_2\text{O}$  (internal reference) in fully hydrated samples. Set at 4.76 ppm at 298 K. Chemical shift decreases by  $\sim 0.1$  ppm for 10 K increase.
- Pure  $\text{H}_3\text{PO}_4$  (internal reference). Set isotropic  $^{31}\text{P}$  resonance at 0 ppm.

*NOTE: With external references, an effort should be made so that the reference volume and NMR tube/rotor are as close as possible to those of the actual sample, to avoid susceptibility effects.*

## *Further reading on NMR referencing*

- Wishart et al, J. Biomol. NMR (1995) 6, 135-140.
- Markley et al, Pure Appl. Chem. 70 (1998) 117.
- Earl & VanderHart, J. Magn. Reson. 48 (1982) 35–54.
- Morcombe & Zilm, Journal of Magnetic Resonance 162 (2003) 479–486.
- Harris et al, Solid State Nuclear Magnetic Resonance 33 (2008) 41–56.

## 2. How to calibrate a variable temperature (VT) unit

Temperature is an important parameter for magnetic alignment, protein rotational diffusion and other dynamics. The variable temperature unit can be easily calibrated by  $^1\text{H}$  NMR.

- Prepare a sample of 100% ethylene glycol (273-416K) or 100% methanol (178-330K ).
- Equilibrate the sample in the spectrometer (~10min)
- Record  $^1\text{H}$  NMR spectra at different VT settings.
- Use the  $^1\text{H}$  chemical shift difference  $\Delta$  (ppm) between methylene/methyl and hydroxyl peaks to determine the actual temperature:

$$T \text{ (K)} = 466.5 - 102 * \Delta \quad \text{(ethylene glycol)}$$

$$T \text{ (K)} = 409 - 36.54 * \Delta - (21.85 * (\Delta^2)) \quad \text{(methanol)}$$

- Plot actual vs nominal (VT) temperatures to determine the correction for a specific VT unit.

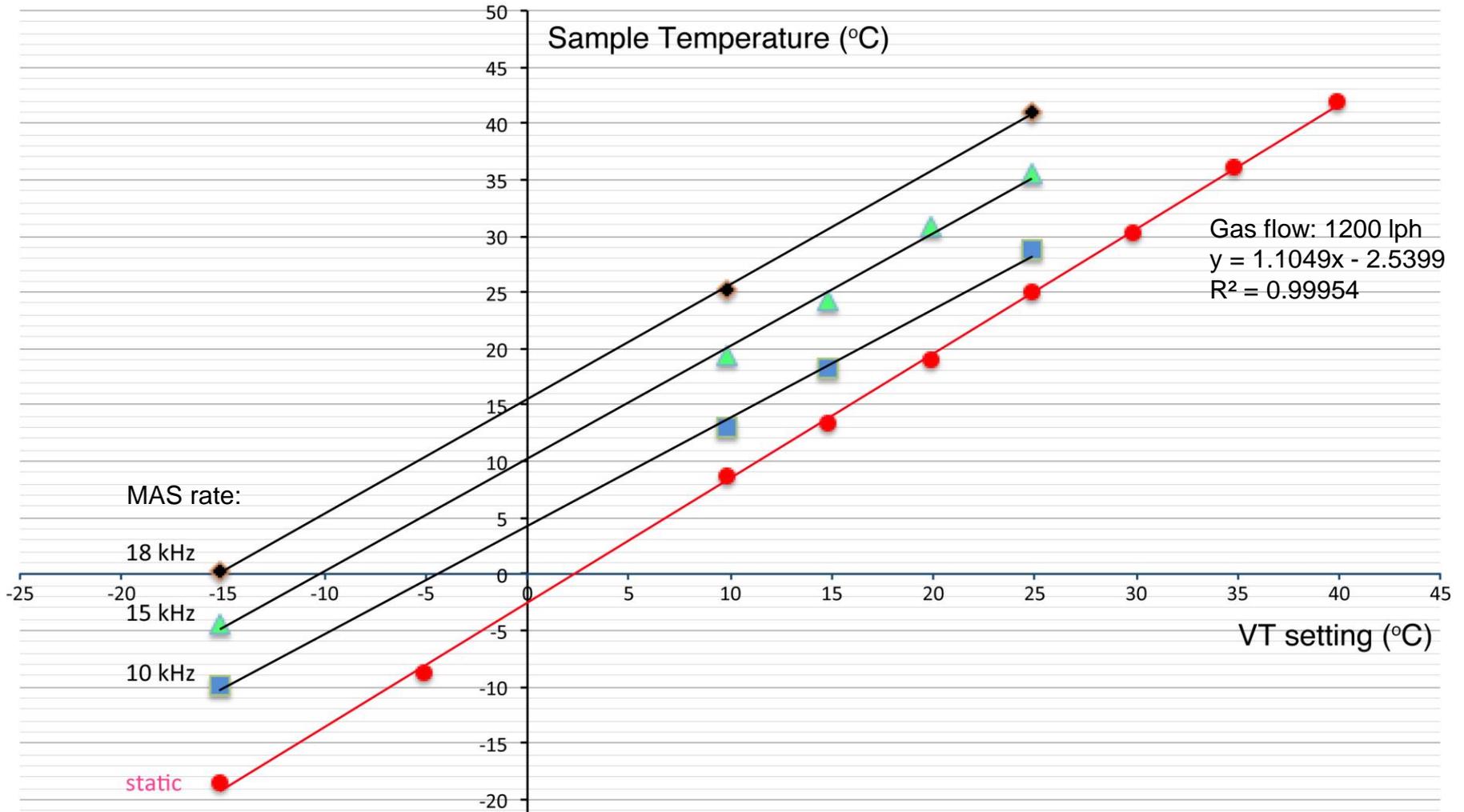
## *Useful resources on temperature standards*

- Kaplan et al, *Anal. Chem.* **1975**, 47, 1703
- Led & Petersen, *J. Magn. Reson.* **1978**, 32, 1 - 17.
- Amman et al, *J. Magn. Reson.* **1982**, 46, 319-321.
- Online NMR temperature calculator:  
[http://www.spectroscopynow.com/userfiles/sepspec/file/specNOW/HTML%20files/NMR\\_temperature\\_measurement.htm](http://www.spectroscopynow.com/userfiles/sepspec/file/specNOW/HTML%20files/NMR_temperature_measurement.htm)

### *3. Frictional heating in hydrated MAS samples*

- Knowledge of the actual sample temperature is important for rotationally aligned (RA) solid state NMR, and in any experiment where protein and/or lipid dynamics is important.
- Spinning liposome or other aqueous protein samples in an MAS experiment increases the inner sample temperature by frictional heating.
- Frictional heating depends on the MAS rate.
- Sample frictional heating can be dependent on chiller setting/air flow rate. A note should be made of these parameters.
- Using VT control, the sample temperature is well equilibrated in 10 minutes.
- The  $^1\text{H}$   $\text{H}_2\text{O}$  resonance inside the sample can be monitored to verify the actual temperature changes in the sample.

# Example: frictional heating in a 3.2 mm Bruker rotor (900MHz Low-E HCN probe, BTRC)



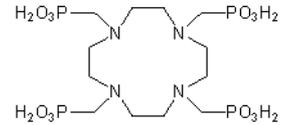
Sample: Ethylene-glycol, verified on H<sub>2</sub>O resonance in biological sample

## *4. Determination of the power deposition in an NMR sample*

- Power deposition during an NMR experiment can be significant in a hydrated biological sample, i.e., a “lossy” sample, and it is strongly probe/coil dependent.
- Lossy samples can be approximated by a 70 mM NaCl aqueous solution.
- $^1\text{H}$  chemical shifts in  $\text{Na}_5[\text{TmDOTP}]$  are used for fast and precise measurements of RF heating (see Zuo et al in references).

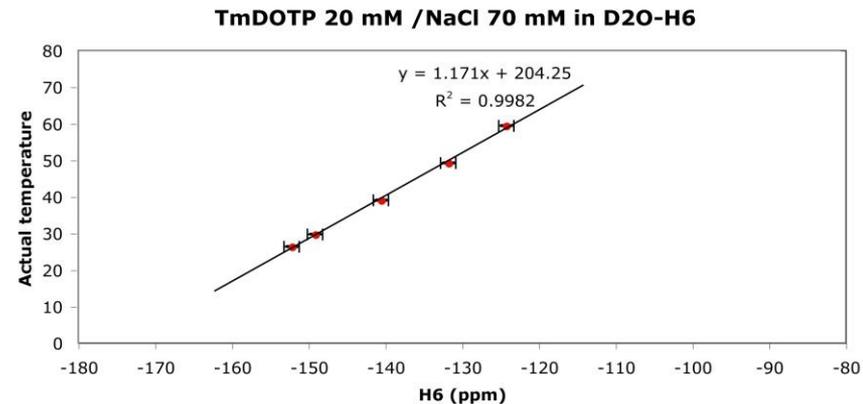
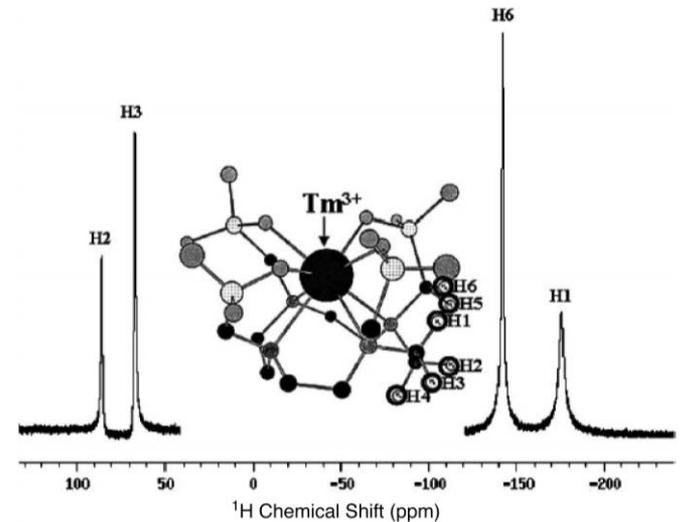
# How to measure RF heating

- Sample: 20 mM Na<sub>5</sub>[TmDOTP] / 70 mM NaCl / D<sub>2</sub>O



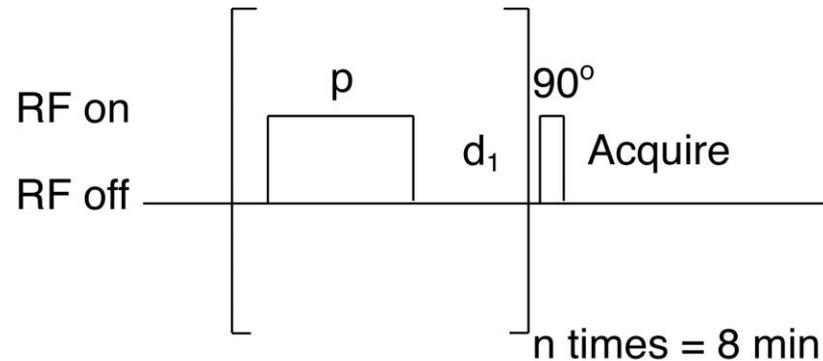
Na<sub>5</sub>[TmDOTP] :

- Biocompatible
  - Versatile: <sup>1</sup>H, <sup>31</sup>P, <sup>23</sup>Na
  - Paramagnetic: short recycle delay ( $d_1 < 500$  ms)
  - Can be also used to measure pH, ions, etc....
  - Cost effective: <100\$/gram from Macrocyclics (TX)
- Calibration curve: measure <sup>1</sup>H shift(s) of Na<sub>5</sub>[TmDOTP] vs sample temperature in a VT-regulated 1-pulse experiment.



# How to measure RF heating

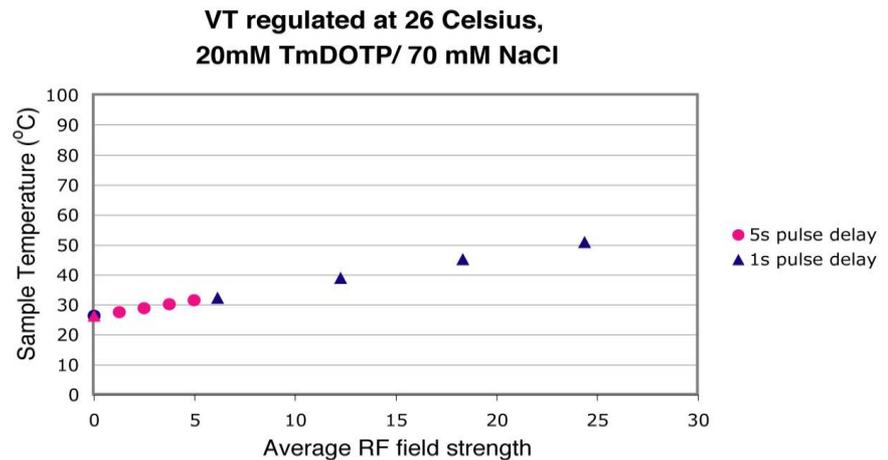
A typical experiment to determine RF heating due to  $^1\text{H}$  irradiation:



“Average” RF =  $B_1^2 \times \text{Duty Factor} =$

$$= B_1^2 \times [\text{time RF on} / \text{time RF off}]$$

where  $B_1$  is in kHz



## *Further reading of RF heating and uses of Na<sub>5</sub>[TmDOTP]*

- Gadian & Robinson, J. Magn. Reson. 34 (1979) 449–455.
- Hoult & Lauterbur, J. Magn. Reson. 34 (1979) 425–433.
- Kelly et al, J. Am. Chem. Soc. 124 (2004) 12013–12019.
- Li et al, Journal of Magnetic Resonance 180 (2006) 51–57
- Zuo et al, Magn Reson Med (1996) 36:955–959
- Zuo et al, J Magn Reson (1998) 133:53–60
  
- Macrocyclics, TX: <https://macrocyclics.com>